

COMMENTARY

Gene therapy to treat inherited and complex retinal degenerative diseases

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These are exciting times in the field of retinal gene therapy, as impressive successes have been achieved by recombinant adeno-associated viral (rAAV) gene delivery of Rpe65 or Rep1 to the retinal pigment epithelium of patients with Lebers congenital amaurosis or choroideremia, respectively.^{1–5} Instead of being left with incurable blindness, patients are reporting restoration of vision. An advantage of these disease targets is that the affected cell type, the retinal pigment epithelium, is amenable to gene therapy since it is naturally phagocytic, resulting in efficient transduction, and is not adversely affected by overexpression of exogenous transgenes. In contrast, the majority of inherited retinal degenerations are due to mutations in photoreceptor-specific transcripts causing primary death of these cells and permanent vision loss.⁶ Novel studies published in *Molecular Therapy—Methods & Clinical Development* address such challenges for photoreceptor gene therapy, including rapid onset of cell death (resulting in pathological changes to the retinal environment) and inefficient transduction of photoreceptors by gene therapy vectors.

Palfi *et al.*⁷ address the latter challenge by demonstrating that rAAV2/rh10 transduces photoreceptors as efficiently as the current gold standard, rAAV2/8.^{8,9} The number of cells transduced and level of transgene expression achieved by Palfi *et al.*, with rAAV2/rh10 is sufficient to provide histological, electrophysiological, and behavioral improvements in a mouse model of autosomal recessive retinitis pigmentosa due to mutations in rhodopsin. Their studies are among the first to show success in the most common form of retinitis pigmentosa.^{10–12} While very exciting and full of promise, the effect is not long-lasting, consistent with results in other studies using rAAV-mediated gene addition to rescue photoreceptors.^{12–19} A decreased rate of degeneration is certainly clinically relevant as it could translate to the addition of years of precious sight in patients. At the same time we need to understand why the efficacy of gene therapy in photoreceptors decreases over time despite addressing the intrinsic deficit (lack of functional protein). This article will discuss two potential explanations: (i) insufficient gene expression within the cells causing intrinsic cell death pathways to still activate; and (ii) changes in the retinal milieu that initiate cell death by extrinsic signaling pathways.

Palfi *et al.*¹¹, both in the recent article in this journal⁷ and in their 2010 paper,¹² achieved a fantastic feat—improvement of structure and function by gene delivery of rhodopsin into the rhodopsin knock-out mouse. Their studies also highlight the challenges of gene augmentation therapy for retinal degeneration. Despite a

significant effort by Palfi *et al.* to increase expression of rhodopsin, the disorganization of the outer segment discs and lower visual function (spatial acuity threshold and electroretinogram) in the treated mice as compared to wild-type controls suggest that normal levels of rhodopsin were still not achieved. Further, the mice still exhibited progressive retinal degeneration. Studies by other groups suggest that increasing gene expression alone may not be sufficient to completely block progression of inherited retinal degenerations. First, increasing gene expression either through repeated injections of rAAV or by use of a more efficient rAAV vector does not overcome this progressive decline in photoreceptors.^{16,17} Second, studies in both dogs and mice show that the therapeutic effects of augmentation gene therapy are more sustained when it is given early, prior to onset of photoreceptor degeneration.^{13,15}

Many inherited retinal degenerations due to deficits in photoreceptor-specific proteins have a very early onset. Therefore, the retinal environment is likely altered prior to intervention by gene therapy. For example, glial reactivity can be detected prior to significant retinal degeneration in multiple models of retinal degeneration.^{20–24} Glial cell reactivity can be beneficial acutely, but can cause oxidative stress and neuroinflammation, leading to neuronal death if the glia remain reactive long-term (for review, see ref. 25). In fact, Palfi *et al.*, detected infiltrates in the rhodopsin knock-out mouse, although they did not assess the level of reactivity of the endogenous glial cells.

However, robust and long-term benefit may be achieved by a combined approach targeting both the intrinsic gene defect and extrinsic signaling (*i.e.*, neuroprotection). This approach has shown success in a mouse model of inherited retinal degeneration.²⁶ Gene delivery of several neuroprotective agents has shown promise in models of inherited retinal degeneration.^{27–34} It is important to note that not all neuroprotective factors are created equal, and all inherited retinal degenerations are not the same. It is critical to have a good understanding of the molecular pathways both activated by the inherited retinal degeneration of interest and inhibited by each potential neuroprotective factor in order to properly pair them. For example, if the photoreceptor degeneration is driven entirely by intrinsic pathways that activate apoptotic cell death, then only those factors that block apoptosis will be effective. In contrast, if autophagy, pyroptosis, or necroptosis are critical, then one needs a factor that will target those pathways. In addition, late stage inherited retinal degenerations, as well as complex retinal degenerations such as glaucoma, diabetic retinopathy, and age-related macular degeneration, are affected by extrinsic factors such as glial reactivity (as mentioned above), oxidative stress, and/or neovascularization, thus requiring factors that will target these processes. Fortunately,

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most neuroprotective factors are pluripotent such that they can address both intrinsic and extrinsic pathways.

Shanab *et al.*³⁵ present a promising neuroprotective gene therapy approach that targets neovascularization in diabetic retinopathy. In the retina, neovascularization most commonly occurs during retinopathy of prematurity and as a common secondary complication to diabetic retinopathy. It can also occur in advanced stages of inherited retinal degenerative diseases, in which the lack of photoreceptors creates a more oxidative environment leading to neovascularization in the retina.³⁶ This group previously demonstrated that an oxidative environment leads to decreased MMP-7 activity resulting in decreased processing of pro-nerve growth factor (NGF) into NGF in both experimental and clinical diabetes.^{37–39} While it is well known that NGF is neuroprotective through activation of TrkA (for review, see ref. 40), they showed that proNGF promoted neuroinflammation and cell death by activation of p75^{NTR}.^{41–43} In their current paper, the authors used virus-mediated (lentivirus) gene delivery of a cleavage resistant proNGF to induce endothelial cell death and counteracted the damage by treatment with a lentivirus carrying shRNA targeting the p75^{NTR}. This approach successfully protected the endothelial cells both *in vivo* and *in vitro*. These results are encouraging and further studies should be performed to test the general applicability of this approach for retinal neovascularization as a result of retinopathy of prematurity, and choroidal neovascularization as a result of age-related macular degeneration or end stage inherited retinal degenerations. There is a need for a new approach for blocking neovascularization since recent studies show that anti-vascular endothelial growth factor therapies are ineffective at blocking neovascularization in a subset of patients.⁴⁴

In summary, two studies now published in *Molecular Therapy—Methods & Clinical Development* highlight the great strides being made in retinal gene therapy and the future of the field. Palfi *et al.* demonstrate great progress in the treatment of retinal degenerations due to lack of functional rhodopsin using a new rAAV serotype. Shanab *et al.* illustrate the utility of virus-mediated gene delivery for understanding the molecular events that underlie disease, in this case neovascularization, and demonstrate a new approach for blocking pathological angiogenesis. This and other neuroprotective approaches will be necessary for treating complex retinal degenerations and may improve outcomes in inherited retinal degenerations. The lack of long-term preservation of vision after gene addition into photoreceptors underscores the need for both gene augmentation and neuroprotection approaches in order to achieve great clinical success. The field is also propelled forward by the development of new serotypes of rAAV that have greater transduction efficiency, faster onset of transgene expression, and target different cell types (*i.e.*, photoreceptors, glia, or ganglion cells). In conclusion, for the field of retinal gene therapy to reach the ultimate goal of treating all forms of blinding disease, multiple groups will need to work together, collaboratively, bringing their expertise on the biology of disease, vector development, and both gene targeted and neuroprotective strategies together to develop synergistic, multipronged gene therapies.

REFERENCES

- Bainbridge, JW, Smith, AJ, Barker, SS, Robbie, S, Henderson, R, Balaggan, K *et al.* (2008). Effect of gene therapy on visual function in Leber's congenital amaurosis. *N Engl J Med* **358**: 2231–2239.
- Jacobson, SG, Acland, GM, Aguirre, GD, Aleman, TS, Schwartz, SB, Cideciyan, AV *et al.* (2006). Safety of recombinant adeno-associated virus type 2-RPE65 vector delivered by ocular subretinal injection. *Mol Ther* **13**: 1074–1084.
- Maguire, AM, Simonelli, F, Pierce, EA, Pugh, EN Jr, Mingozzi, F, Bennicelli, J *et al.* (2008). Safety and efficacy of gene transfer for Leber's congenital amaurosis. *N Engl J Med* **358**: 2240–2248.
- Simonelli, F, Maguire, AM, Testa, F, Pierce, EA, Mingozzi, F, Bennicelli, JL *et al.* (2010). Gene therapy for Leber's congenital amaurosis is safe and effective through 1.5 years after vector administration. *Mol Ther* **18**: 643–650.
- Jacobson, SG, Cideciyan, AV, Ratnakaram, R, Heon, E, Schwartz, SB, Roman, AJ *et al.* (2012). Gene therapy for leber congenital amaurosis caused by RPE65 mutations: safety and efficacy in 15 children and adults followed up to 3 years. *Arch Ophthalmol* **130**: 9–24.
- RetNet. 1996–2009. <http://www.sph.uth.tmc.edu/retnet/>.
- Palfi, A, Chadderton, N, O'Reilly, M, Nagel-Wolfrum, K, Wolfrum, U, Bennett, J *et al.* (2015). Efficient gene delivery to photoreceptors using AAV2/rh10 and rescue of the Rho-/- mouse. *Mol Ther Methods Clin Dev* **2**: 15016.
- Allocca, M, Mussolino, C, Garcia-Hoyos, M, Sanges, D, Iodice, C, Petrillo, M *et al.* (2007). Novel adeno-associated virus serotypes efficiently transduce murine photoreceptors. *J Virol* **81**: 11372–11380.
- Natkunarahaj, M, Trittibach, P, McIntosh, J, Duran, Y, Barker, SE, Smith, AJ *et al.* (2008). Assessment of ocular transduction using single-stranded and self-complementary recombinant adeno-associated virus serotype 2/8. *Gene Ther* **15**: 463–467.
- Mao, H, Gorbatyuk, MS, Rossmiller, B, Hauswirth, WW and Lewin, AS (2012). Long-term rescue of retinal structure and function by rhodopsin RNA replacement with a single adeno-associated viral vector in P23H RHO transgenic mice. *Hum Gene Ther* **23**: 356–366.
- Mao, H, James, T Jr, Schwein, A, Shabashvili, AE, Hauswirth, WW, Gorbatyuk, MS *et al.* (2011). AAV delivery of wild-type rhodopsin preserves retinal function in a mouse model of autosomal dominant retinitis pigmentosa. *Hum Gene Ther* **22**: 567–575.
- Palfi, A, Millington-Ward, S, Chadderton, N, O'Reilly, M, Goldmann, T, Humphries, MM *et al.* (2010). Adeno-associated virus-mediated rhodopsin replacement provides therapeutic benefit in mice with a targeted disruption of the rhodopsin gene. *Hum Gene Ther* **21**: 311–323.
- Cideciyan, AV, Jacobson, SG, Beltran, WA, Sumaroka, A, Swider, M, Iwabe, S *et al.* (2013). Human retinal gene therapy for Leber congenital amaurosis shows advancing retinal degeneration despite enduring visual improvement. *Proc Natl Acad Sci USA* **110**: E517–E525.
- Pawlyk, BS, Smith, AJ, Buch, PK, Adamian, M, Hong, DH, Sandberg, MA *et al.* (2005). Gene replacement therapy rescues photoreceptor degeneration in a murine model of Leber congenital amaurosis lacking RPGRIP. *Invest Ophthalmol Vis Sci* **46**: 3039–3045.
- Sarra, GM, Stephens, C, de Alwis, M, Bainbridge, JW, Smith, AJ, Thrasher, AJ *et al.* (2001). Gene replacement therapy in the retinal degeneration slow (rds) mouse: the effect on retinal degeneration following partial transduction of the retina. *Hum Mol Genet* **10**: 2353–2361.
- Schlichtenbrede, FC, da Cruz, L, Stephens, C, Smith, AJ, Georgiadis, A, Thrasher, AJ *et al.* (2003). Long-term evaluation of retinal function in Prph2Rd2/Rd2 mice following AAV-mediated gene replacement therapy. *J Gene Med* **5**: 757–764.
- Allocca, M, Manfredi, A, Iodice, C, Di Vicino, U and Auricchio, A (2011). AAV-mediated gene replacement, either alone or in combination with physical and pharmacological agents, results in partial and transient protection from photoreceptor degeneration associated with betaPDE deficiency. *Invest Ophthalmol Vis Sci* **52**: 5713–5719.
- Koch, S, Sothilingam, V, Garcia Garrido, M, Tanimoto, N, Becirovic, E, Koch, F *et al.* (2012). Gene therapy restores vision and delays degeneration in the CNGB1(-/-) mouse model of retinitis pigmentosa. *Hum Mol Genet* **21**: 4486–4496.
- Smith, AJ, Schlichtenbrede, FC, Tschernutter, M, Bainbridge, JW, Thrasher, AJ and Ali, RR (2003). AAV-Mediated gene transfer slows photoreceptor loss in the RCS rat model of retinitis pigmentosa. *Mol Ther* **8**: 188–195.
- Zeng, HY, Zhu, XA, Zhang, C, Yang, LP, Wu, LM and Tso, MO (2005). Identification of sequential events and factors associated with microglial activation, migration, and cytotoxicity in retinal degeneration in rd mice. *Invest Ophthalmol Vis Sci* **46**: 2992–2999.
- Guo, C, Otani, A, Oishi, A, Kojima, H, Makiyama, Y, Nakagawa, S *et al.* (2012). Knockout of ccr2 alleviates photoreceptor cell death in a model of retinitis pigmentosa. *Exp Eye Res* **104**: 39–47.
- Noailles, A, Fernández-Sánchez, L, Lax, P and Cuenca, N (2014). Microglia activation in a model of retinal degeneration and TUDCA neuroprotective effects. *J Neuroinflammation* **11**: 186.
- Eisenfeld, AJ, Bunt-Milam, AH and Sarthy, PV (1984). Müller cell expression of glial fibrillary acidic protein after genetic and experimental photoreceptor degeneration in the rat retina. *Invest Ophthalmol Vis Sci* **25**: 1321–1328.
- de Kozak, Y, Cotinet, A, Goureau, O, Hicks, D and Thillaye-Goldenberg, B (1997). Tumor necrosis factor and nitric oxide production by resident retinal glial cells from rats presenting hereditary retinal degeneration. *Ocul Immunol Inflamm* **5**: 85–94.
- Karlstetter, M, Scholz, R, Rutar, M, Wong, WT, Provis, JM and Langmann, T (2015). Retinal microglia: just bystander or target for therapy? *Prog Retin Eye Res* **45**: 30–57.
- Buch, PK, MacLaren, RE, Durán, Y, Balaggan, KS, MacNeil, A, Schlichtenbrede, FC *et al.* (2006). In contrast to AAV-mediated Cntf expression, AAV-mediated Gdnf expression enhances gene replacement therapy in rodent models of retinal degeneration. *Mol Ther* **14**: 700–709.

27. Rex, TS, Allocca, M, Domenici, L, Surace, EM, Maguire, AM, Lyubarsky, A *et al.* (2004). Systemic but not intraocular Epo gene transfer protects the retina from light- and genetic-induced degeneration. *Mol Ther* **10**: 855–861.
28. Rex, TS, Wong, Y, Kodali, K and Merry, S (2009). Neuroprotection of photoreceptors by direct delivery of erythropoietin to the retina of the retinal degeneration slow mouse. *Exp Eye Res* **89**: 735–740.
29. Sullivan, T, Kodali, K and Rex, TS (2011). Systemic gene delivery protects the photoreceptors in the retinal degeneration slow mouse. *Neurochem Res* **36**: 613–618.
30. Liang, FQ, Dejneka, NS, Cohen, DR, Krasnoperova, NV, Lem, J, Maguire, AM *et al.* (2001). AAV-mediated delivery of ciliary neurotrophic factor prolongs photoreceptor survival in the rhodopsin knockout mouse. *Mol Ther* **3**: 241–248.
31. Dalkara, D, Kolstad, KD, Guerin, KI, Hoffmann, NV, Visel, M, Klimczak, RR *et al.* (2011). AAV mediated GDNF secretion from retinal glia slows down retinal degeneration in a rat model of retinitis pigmentosa. *Mol Ther* **19**: 1602–1608.
32. Gregory-Evans, K, Chang, F, Hodges, MD and Gregory-Evans, CY (2009). Ex vivo gene therapy using intravitreal injection of GDNF-secreting mouse embryonic stem cells in a rat model of retinal degeneration. *Mol Vis* **15**: 962–973.
33. Murakami, Y, Ikeda, Y, Yonemitsu, Y, Onimaru, M, Nakagawa, K, Kohno, R *et al.* (2008). Inhibition of nuclear translocation of apoptosis-inducing factor is an essential mechanism of the neuroprotective activity of pigment epithelium-derived factor in a rat model of retinal degeneration. *Am J Pathol* **173**: 1326–1338.
34. Leonard, KC, Petrin, D, Coupland, SG, Baker, AN, Leonard, BC, LaCasse, EC *et al.* (2007). XIAP protection of photoreceptors in animal models of retinitis pigmentosa. *PLoS ONE* **2**: e314.
35. Shanab, AY, Mysona, BA, Matragoon, S and El-Remessy, AB (2015). Silencing p75NTR prevents proNGF-induced endothelial cell death and development of acellular capillaries in rat retina. *Mol Ther Methods Clin Dev* **2**: 15013.
36. Marano, F, Deutman, AF, Leys, A and Aandeker, AL (2000). Hereditary retinal dystrophies and choroidal neovascularization. *Graefes Arch Clin Exp Ophthalmol* **238**: 760–764.
37. Ali, TK, Matragoon, S, Pillai, BA, Liou, GI and El-Remessy, AB (2008). Peroxynitrite mediates retinal neurodegeneration by inhibiting nerve growth factor survival signaling in experimental and human diabetes. *Diabetes* **57**: 889–898.
38. Al-Gayyar, MM, Abdelsaid, MA, Matragoon, S, Pillai, BA and El-Remessy, AB (2011). Thioredoxin interacting protein is a novel mediator of retinal inflammation and neurotoxicity. *Br J Pharmacol* **164**: 170–180.
39. Ali, TK, Al-Gayyar, MM, Matragoon, S, Pillai, BA, Abdelsaid, MA, Nussbaum, JJ *et al.* (2011). Diabetes-induced peroxynitrite impairs the balance of pro-nerve growth factor and nerve growth factor, and causes neurovascular injury. *Diabetologia* **54**: 657–668.
40. Tabakman, R, Lecht, S, Sephanova, S, Arien-Zakay, H and Lazarovici, P (2004). Interactions between the cells of the immune and nervous system: neurotrophins as neuroprotection mediators in CNS injury. *Prog Brain Res* **146**: 387–401.
41. Al-Gayyar, MM, Mysona, BA, Matragoon, S, Abdelsaid, MA, El-Azab, MF, Shanab, AY *et al.* (2013). Diabetes and overexpression of proNGF cause retinal neurodegeneration via activation of RhoA pathway. *PLoS ONE* **8**: e54692.
42. Mysona, BA, Al-Gayyar, MM, Matragoon, S, Abdelsaid, MA, El-Azab, MF, Saragovi, HU *et al.* (2013). Modulation of p75(NTR) prevents diabetes- and proNGF-induced retinal inflammation and blood-retina barrier breakdown in mice and rats. *Diabetologia* **56**: 2329–2339.
43. Matragoon, S, Al-Gayyar, MM, Mysona, BA, Abdelsaid, MA, Pillai, BA, Neet, KE *et al.* (2012). Electroporation-mediated gene delivery of cleavage-resistant pro-nerve growth factor causes retinal neuro- and vascular degeneration. *Mol Vis* **18**: 2993–3003.
44. Dedania, VS, Grob, S, Zhang, K and Bakri, SJ (2015). Pharmacogenomics of response to anti-VEGF therapy in exudative age-related macular degeneration. *Retina (Philadelphia, Pa)* **35**: 381–391.



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